

VII. CLAIMING

What is claimed is:

1. The procedure for cloning human SMN gene based on the reverse transcription (RT) and the polymerase chain reaction (PCR) using the synthesized oligonucleotides (SEQ ID NO. 1) for RT, and (SEQ ID NO. 2) and (SEQ ID NO. 3) respectively for PCR, comprising:

- Isolating RNA;

- Performing RT reaction using the synthesized oligonucleotide

5' TGGCAGACTTAC 3' (SEQ ID NO. 1) under the following conditions: 90°C

for 2 minutes; 0°C for 1 minute; 25°C for 10 minutes; 42°C for 45 minutes;

- Performing PCR reaction using the synthesized oligonucleotides

5' ATGGCGATGAGCAGCGG 3' (SEQ ID NO. 2) and

5' TTAATTTAAGGAATGTGAGCAC 3' (SEQ ID NO. 3) under the following conditions: Denaturing at 94°C for 1 minute; annealing at 55°C for 2 minutes;

elongating at 72°C for 1 minute each cycle, for 35 cycles.

2. The procedure for the construction of expression plasmids using the pFastBacTM HTb and the pBlueBacHis2 A transfer vectors for the purpose of obtaining human SMN protein in insect cells, comprising:

- 2.1. Using the pFastBacTM HTb vector:

- Digesting the pFastBacTM HTb vector with BamHI and XhoI followed by dephosphorylation with calf intestinal alkaline phosphatase;

- Digesting the vector (1) pCR^R II/SMN-cDNA with BamHI and XhoI and isolating the resulting fragment containing the cDNA coding sequences of SMN protein, SMN- cDNA;
- Ligating the SMN-cDNA fragment to the pFastBacTM HTb vector and introducing the ligation product in INV α F' E. Coli strain;
- Screening for inserts based on the presence of white colonies, as a result of which the vector (2) pFastBacTM HTb/SMN-cDNA is selected;
- Introducing the vector (2) in DH10BacTM E. Coli competent cells;
- Screening for recombinant bacmids in DH10BacTM E. Coli using blue-white color selection, then verifying the presence of SMN-cDNA's insert in the recombinant bacmids by PCR amplification using the M13 forward (-40) and M13 reverse primers, as a result of which the recombinant bacmid (3) is selected;

2.2. Using the pBlueBacHis2 A vector:

- Digesting the pBlueBacHis2 A vector with BamHI and XhoI followed by dephosphorylation with calf intestinal alkaline phosphatase;
- Digesting the vector (2) pFastBacTM HTb/SMN-cDNA with BamHI and XhoI and isolating the resulting fragment containing the cDNA coding sequences of SMN protein, SMN-cDNA;
- Ligating the SMN-cDNA fragment to the pBlueBacHis2 A vector and introducing the ligation product in INV α F' E. Coli strain;
- Screening for inserts using blue-white color selection, as a result of which the vector (4) pBlueBacHis2 A/SMN-cDNA is selected.

3. The procedure for the construction of expression plasmids using the pET-28a (+) transfer vector for the purpose of obtaining human SMN protein in bacteria, comprising:

- Digesting the pET-28a (+) vector with BamHI and XhoI followed by dephosphorylation with calf intestinal alkaline phosphatase;

5 - Digesting the vector (2) pFastBacTM HTb/SMN-cDNA with BamHI and XhoI and isolating the resulting fragment containing the cDNA coding sequences of SMN protein, SMN-cDNA;

- Ligating the SMN-cDNA fragment to the pET-28a (+) vector and introducing the ligation product in INV α F' E. Coli strain;

10 - Screening for inserts based on the presence of white colonies, as a result of which the vector (5) pET-28a (+)/SMN-cDNA is selected.

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VIII. REFERENCES

1. Roberts, D.F.; Chavez, J.; Court, S.D.M. The genetic component in child mortality. Arch. Dis. Child **45**, (1970) 33-38.
2. Pearn, J. The gene frequency of acute Werdnig-Hoffmann disease (SMA type I). A total
5 population survey in North East England. J. Med. Genet. **10**, (1973) 260-265.
3. Pearn, J. Incidence, prevalence, and gene frequency studies of chronic childhood spinal
muscular atrophy. J. Med. Genet. **15**, (1978) 409-413.
4. Czeizel, A.; Hamula, J. A Hungarian study on Werdnig-Hoffmann disease. J. Med.
Genet. **26**, (1989) 761-763.
- 10 5. Munsat, T.L. Workshop report: International SMA collaboration. Neuromusc. Disord. **1**,
(1991) 81.
6. Werdnig, G. Die fruhsinfantile progressive spinal atrophie. Arch. Psych. **26**, (1894) 706-
744.
7. Hoffmann, J. Uber die hereditare progressive spinal muskelatrophie im Kindesalter.
15 Munchen Med. Wschr. **47**, (1900) 1649-1651.
8. Kugelberg, E.; Welander, L. Heredo-familial juvenile muscular atrophy simulating
muscular dystrophy. Arch. Neurol. Psych. **75**, (1956) 500-509.
9. Brzustowicz, L.M.; Lehner, T.; Castilla, L.H.; et al. Genetic mapping of chronic
childhood-onset spinal muscular atrophy to chromosome 5q 11.2- q 13.3. Nature **344**,
20 (1990) 540-541.
10. Gilliam, T.C.; Brzustowicz, L.M.; Castilla, L.H.; et al. Genetic homogeneity between
acute and chronic forms of spinal muscular atrophy. Nature **345**, (1990) 823-825.

11. Melki, J.; Abdelhak, S.; Sheth, P.; et al. Gene for proximal spinal muscular atrophies maps to chromosome 5q. Nature **344**, (1990) 767-768.
12. Melki, J.; Sheth, P.; Abdelhak, S.; et al. Mapping of acute (type I) spinal muscular atrophy to chromosome 5q12-q14. Lancet **336**, (1990) 271-273.
- 5 13. Melki, J.; Lefebvre, S.; Burglen, L.; et al. De novo and inherited deletions of the 5q13 region in spinal muscular atrophies. Science **264**, (1994) 1474-1477.
14. Lefebvre, S.; Burglen, L.; Reboullet, S.; et al. Identification and characterization of a spinal muscular atrophy-determining gene. Cell **80**, (1995) 155-165.
15. Roy, N.; Mahadevan, M.; Mclean, M.; et al. The gene for neuronal apoptosis inhibitory
10 protein is partially deleted in individuals with spinal muscular atrophy. Cell **80**, (1995) 167-178.
16. Fischer, U.; Liu, Q.; Dreyfuss, G. The SMN-SIP-1 complex has an essential role in spliceosomal snRNP biosynthesis. Cell **90**, (1997) 1023-1029.
17. Liu, Q.; Fischer, U.; Wang, F.; Dreyfuss, G. The spinal muscular atrophy disease gene
15 product, SMN, and its associated protein SIP-1 are in complex with spliceosomal snRNP proteins. Cell **90**, (1997) 1013-1021.
18. Pellizzoni, L.; Kataoka, N.; Charroux, B.; Dreyfuss, G. A novel function for SMN, the spinal muscular atrophy disease gene product, in pre-mRNA splicing. Cell **95**, (1998) 615-624.
- 20 19. Schrank, B.; Gotz, R.; Gunneron, J.; Ure, J.; Toyka, K.; Smith, A.; Sendtner, M. Inactivation of the survival motor neuron gene, a candidate gene for human spinal muscular atrophy, leads to massive cell death in early mouse embryos. Proc. Natl. Acad. Sci. USA **94**, (1997) 9920-9925.

20. Burlet, P.; Huber, C.; Bertrand, S.; Ludosky, M. A.; Zwaenepoel, I.; Clermont, O.; Roume, J.; Delezoide, A.L.; Cartaud, J.; Munnich, A.; Lefebvre, S. The distribution of SMN protein complex in human fetal tissues and its alteration in spinal muscular atrophy. Hum. Mol. Genet. **7**, (1998) 1927-1933.
- 5 21. Lefebvre, S.; Burlet, P.; Liu, Q.; Bertrand, S.; Clermont, O.; Munnich, A.; Dreyfuss, G.; Melki, J. Correlation between severity and SMN protein level in spinal muscular atrophy. Nature Genet. **16**, (1997) 66-70.
22. Monani, U.R.; Sendtner, M.; Covert, D.D.; Parsons, D.W.; Andreassi, C.; Le, T.T.; Jablonka, S.; Schrank, B.; Rossoll, W.; Prior, T.W.; Morris, G.E.; Burghes, A.H.M. The human centromeric survival motor neuron gene (SMN 2) rescues embryonic lethality in *Smm*^{-/-} mice and results in a mouse with spinal muscular atrophy. Hum. Mol. Genet. **9**, (2000) 333-339.
- 10 23. Jablonka, S.; Schrank, B.; Kralewski, M.; Rossoll, W.; Sendtner, M. Reduced survival motor neuron (Smn) gene dose in mice leads to motor neuron degeneration: an animal model for spinal muscular atrophy type III. Hum. Mol. Genet. **9**, (2000) 341-346.
- 15 24. Li, H.M.H.; Chang, J.G.; Jong, Y.J.; Wu, M.H.; Wang, N.M.; Tsai, C.H.; Li, H. A mouse model for spinal muscular atrophy. Nature Genet. **24**, (2000) 66-70.
25. Sambrook, J.; Fritsch, E.F.; Maniatis, T. Extraction, purification and analysis of messenger RNA from eukaryotic cells. In Molecular Cloning, a Laboratory Manual, 2nd Ed. Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York (1989) 7.28-7.52.
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26. Saiki, R.K.; Scharf, S.; Faloona, F; Mullis, K.B.; Horn, C.T.; Erlich, H.A.; Arnheim, N. Amplification of β -globine genomic sequences and restriction site and analysis for diagnosis of sickle cell anemia. Science **230** (1985) 1350-1354.
27. Kawasaki, E.S.; Wang, A.M. Detection of gene expression. In PCR Technology, Erlich, H.A., Ed. Stockton: New York (1989) 89-97.
28. Arnold, A.S.; Gueye, M.; Ronde, P.; Warter, J.M.; Poindron, P.; Gies, J.P. Construction of a plasmid containing human SMN, the SMA determining gene, coupled to EGFP. Plasmid **47**, (2002) 79-87.